

WHITEFLY RESISTANCE TO INSECTICIDES IN ARIZONA: SUMMARY OF 2002 AND 2003 RESULTS

Timothy J. Dennehy, Benjamin A. DeGain, Virginia S. Harpold, and Sarah A. Brink

Department of Entomology, The University of Arizona, Tucson, AZ

Robert L. Nichols

Cotton Incorporated, Cary, NC

SUMMARY

The development of pest resistance poses a constant threat to successful management of sticky cotton resulting from inadequate control of Bemisia whiteflies. A three-stage resistance management program was implemented in Arizona cotton following severe whitefly resistance in 1995. This program has been highly successful for eight years. Success has been fostered by intensive investments into improved whitefly sampling and treatment decisions, coupled with conservation of natural enemies. This latter component has hinged on limited, strategic use of two insect growth regulators in cotton, use of the neonicotinoid insecticide, imidacloprid, in vegetables and melons, and tactical deployment of non-pyrethroid and pyrethroid chemicals. Statewide monitoring of whitefly resistance to insecticides in cotton, melons and greenhouse crops has permitted annual assessments of the status of whitefly resistance management in Arizona. In this paper we summarize susceptibility of whitefly collections made in cotton in the 2002 and 2003 seasons and discuss longer term trends in resistance development. No major problems regarding field performance of insecticides against whiteflies were observed or reported in 2002 or 2003. However, monitoring confirmed the presence of whiteflies with decreased susceptibility to pyriproxyfen (Knack[®]) and showed that they could be detected in all cotton-producing areas of the state. Susceptibility to buprofezin (Courier[®]/Applaud[®]) has not changed significantly since 1997. Mean susceptibility to synergized pyrethroids (e.g., Danitol[®] + Orthene[®]) has increased strikingly on a statewide basis since 1995. However, 50 and 25% of cotton fields sampled in 2002 and 2003, respectively, had resistance levels that have been associated with inadequate field performance of synergized pyrethroid treatments. Whiteflies from throughout Arizona were highly susceptible to imidacloprid (Admire[®]/Provado[®]) and two other neonicotinoid insecticides, acetamiprid (Intruder[®]) and thiamethoxam (Actara[®]/Centric[®]/Platinum[®]).

INTRODUCTION

The neonicotinoid insecticide, imidacloprid (Admire[®]/Provado[®]), and the growth-regulating insecticides (IGRs), buprofezin (Courier[®]/Applaud[®]) and pyriproxyfen (Knack[®]), serve critical roles in controlling whiteflies (*Bemisia tabaci*, a.k.a. *Bemisia argentifolii*) in the Arizona's low desert agricultural ecosystems (Dennehy and Williams 1997, Kerns and Palumbo 1995), as well as in other arid regions of the world (Denholm et al. 1998). Imidacloprid has provided successful season-long whitefly control in Arizona vegetables

and melons since 1993, and has been used on a high proportion of these crops since its introduction (Palumbo 2003). The IGRs, buprofezin and pyriproxyfen, were introduced to Arizona cotton in 1996, after resistance to synthetic pyrethroids and other conventional insecticides reached crisis proportions in 1995 (Dennehy et al. 1996). Buprofezin and pyriproxyfen have provided the foundation for a successful resistance management strategy, their use against whiteflies in cotton being limited to once per season for each. Since 1995,

insecticide treatments in Arizona cotton have declined to averages of less than two treatments per year (Agnew and Baker 2001, Shanley and Baker 2002, 2003). This represents a dramatic change from 1995 when producers were making 6 to 12 insecticide treatments per acre of cotton. Thus, intensive investments into improved monitoring and management of whiteflies (Ellsworth et al. 1996, Ellsworth and Martinez-Carillo 2001), coupled with availability of highly effective, selective insecticides, have greatly reduced the costs of controlling whiteflies in cotton. Sustaining successful whitefly management in Arizona will hinge foremost on avoiding resistance to neonicotinoid insecticides used in melons and vegetables and to insect growth regulators used in cotton.

Whiteflies have been shown to be capable of developing resistance to imidacloprid, pyriproxyfen, and buprofezin under both laboratory and field exposure conditions. An up-to 82-fold resistance to imidacloprid was selected by Prabhaker et al. (1997) under laboratory conditions. Control failures with whiteflies have been reported in greenhouses in Spain to imidacloprid (Denholm et al. 1998). Whitefly resistance has been documented to buprofezin and pyriproxyfen in Israel (Horowitz and Ishaaya 1994, Horowitz et al. 1994, 1999, 2002). Whitefly resistance is monitored yearly in Arizona in order to evaluate the appropriateness of resistance management recommendations and to identify resistance problems and potential solutions before they result in severe economic losses to growers. In this paper we report the results of monitoring of whitefly resistance conducted throughout Arizona cotton in the 2002 and 2003 seasons.

MATERIALS AND METHODS

Whitefly Collections

More than 80 cotton fields, 30 melon fields and 30 retail nurseries were sampled for whiteflies during 2002 and 2003. Herein, we report only results of collections from cotton. Our objective was to obtain a minimum of 1000 individuals from each location. Low whitefly densities, field treatments

with insecticides, and predation/parasitism prevented successful rearing of some collections. Whitefly cultures were successfully established (Table 1a) from 12 Arizona cotton sites in 2002 and from 17 Arizona cotton sites and one California cotton site in 2003 (Table 1b).

Adult whiteflies were collected in modified plastic vials by vacuuming plant foliage with a Makita® Cordless Vacuum (Model 4071D). Samples were chilled and transported to the laboratory in Tucson within eight hours. Samples were released into cages containing several cotton plants, *Gossypium hirsutum* L. (var. DPL-50), at the five to seven true-leaf stages. Adult whiteflies were assayed approximately 12-36 hours after field collection. A reference population, Somerton'93, was maintained since its collection in Arizona in 1993 on cotton plants in laboratory cages without insecticide exposure. Somerton'93 was tested repeatedly each year to provide an internal control of bioassay methodology.

Bioassay Methods

Bioassays were conducted with six insecticides on each culture, when numbers of whiteflies in cultures permitted (Table 1b). Bioassay methods for pyriproxyfen and buprofezin were described by Li et al. (2000, 2003). The residual leaf-disk bioassay used for fenpropathrin + acephate mixtures was described by Dennehy and William (1997). All three neonicotinoid insecticides, imidacloprid, thiamethoxam, and acetamiprid, were testing using leaf disk bioassays (Li et al. 2000) and conditions noted in Table 1c.

Data analyses

Mean mortality observed with all concentrations of the six insecticides evaluated was corrected for control mortality using Abbott's correction (Abbott 1925). Statistical differences in population responses within and between years were evaluated by analysis of variance (ANOVA, Tukey-Kramer HSD test) and non-parametric tests using the JMP-IN statistical analysis program (SAS Institute 2000). Mortality data were subjected to arcsine transformation before analysis. When appropriate, probit analyses of the concentration-dependent

mortality were undertaken using POLO-PC (LeOra Software, 1987) to generate lethal concentration statistics.

RESULTS AND DISCUSSION

Pyriproxyfen (Knack)

Statewide Averages 1996 to 2003. During the first three years pyriproxyfen was used in Arizona, 1996-98, statewide averages of mortality in discriminating concentration bioassays of 0.1 µg/ml pyriproxyfen were $\geq 99.6\%$ (Figure 1a). Approximately 5.5% of whiteflies collected from cotton survived these treatments in 2002; the grand mean of corrected mortality was 94.5% (Figure 1a). In 2003; 15% of whiteflies survived treatments of 0.1 µg/ml pyriproxyfen; the grand mean of corrected mortality was 85.3% (Figure 1a). Changes in mortality in bioassays of 0.01 µg pyriproxyfen/ml were even more dramatic over this same period. Grand mean mortality was $> 80\%$ from 1996 to 1998. This fell to $< 40\%$ in 2003 (Figure 1a).

Resistance Levels at Individual Field Sites. To contrast the frequency of whiteflies with significantly reduced susceptibility to pyriproxyfen at individual sites tested, we have used 2.0% survivors of 0.1 µg/ml pyriproxyfen bioassays as a critical frequency. None of the 48 cultures evaluated from 1996 to 1998 had $\geq 2.0\%$ whiteflies surviving 0.1 µg/ml pyriproxyfen bioassays (Figure 1a). Indeed, as detailed above, survivors of 0.1 µg/ml pyriproxyfen bioassays were very rare for the first three years that pyriproxyfen was used, and constituted $\leq 0.4\%$ of whiteflies tested each year.

Eight of the 12 cotton sites tested in 2002 (67%) had $>2.0\%$ corrected survivorship of 0.1 µg/ml pyriproxyfen (Figure 1b). In 2003, 14 of the 18 sites evaluated (78%) had $>2.0\%$ corrected survivorship of 0.1 µg/ml pyriproxyfen (Figure 1b). One collection made in 2002 and three made in 2003 had greater than 20% corrected survival of 0.1 µg/ml pyriproxyfen. The most resistant collection both years came from the Maricopa Agricultural Center. In 1996, the first year of use of Knack in cotton, a whitefly collection from the Maricopa

Agricultural Center was shown to have an LC_{50} of 0.0033 µg/ml pyriproxyfen (Simmons et al. 1997). Probit analysis of responses of 2002 and 2003 collections from this same location yielded LC_{50} estimates of 0.045 and 0.16, respectively (Figure 1c). On this basis, the Maricopa Agricultural Center whiteflies were 13-fold less susceptible in 2002 and 48-fold less susceptible in 2003 than in 1996.

Selection for a Highly Pyriproxyfen-Resistant Laboratory Strain. We have intensively selected Arizona whiteflies for resistance to pyriproxyfen in the laboratory each year since 1996. We were unsuccessful at isolating resistant strains until 2002. In that year a collection from Queen Creek (Figure 1b, Table 1a) had approximately 10% survivors of 0.1 µg/ml pyriproxyfen when bioassayed directly from the field. Over the subsequent six months we exposed a sub-strain twice to treatments of 0.1 µg/ml pyriproxyfen. Both selected and non-selected Queen Creek sub-strains were reared and tested simultaneously. The outcome of two exposures to pyriproxyfen in the laboratory was a 1000-fold increase in resistance to pyriproxyfen (Figure 1d). In April of 2003, the selected strain had $< 30\%$ mortality in bioassays of 0.1 µg/ml pyriproxyfen and $< 90\%$ mortality in bioassays of 10 µg/ml pyriproxyfen. We have not detected field populations expressing this high level of resistance.

Conclusions Regarding Resistance to Pyriproxyfen. The Arizona Whitefly Resistance Working Group continues to recommend the use of either pyriproxyfen (Knack) or buprofezin (Courier/Applaud) as the first treatments against whiteflies in cotton. It is clear from our findings that whiteflies in some areas of Arizona are substantially less susceptible to pyriproxyfen than they were previously. However, this finding does not mean that Knack has failed under field conditions or that failure is imminent. As already stated, we know of no reports of field failures in Arizona cotton. We cannot predict future changes in susceptibility with accuracy. It is possible that the decreases in susceptibility that we documented during the past three years (Figure 1a) may be reversed in the future. Monitoring of susceptibility of Arizona whiteflies will be continued throughout

the state in order to permit yearly assessments of this situation. In addition, we will continue to work closely with the Arizona Cotton Growers Association, the Arizona Cotton Research and Protection Council, The Arizona Whitefly Working Group, and Valent USA Corporation, to provide Arizona cotton growers with the best information and strategies possible for responding to future developments.

Typically agricultural producers must experience expensive field failures of insecticides before attention is given to resistance problems. Once products fail in the field one or more years of research is often required to document the problem and to formulate strategies to manage resistance. This outcome has negative financial implications for producers, especially as it pertains to stickiness of cotton resulting from whiteflies. Multiple years of discounted cotton prices can result from a single year in which buyers experience stickiness in a region's cotton (Ellsworth et al. 1999). By monitoring resistance pro-actively, i.e., prior to the onset of field problems, we strive to minimize resistance-related costs to producers. Thus, our reason for reporting the early stages of resistance to pyriproxyfen at this time is to increase producer awareness so that they will be most likely to assist us in identifying future developments. The sooner that we are able to detect new resistance problems in the field, the more likely we will be to have the needed time to isolate and manage the problem.

Buprofezin (Courier/Applaud)

Mean susceptibility of Arizona whiteflies to buprofezin in 2002 and 2003 is illustrated in Figure 2. We previously reported a small but statistically significant reduction in mortality observed in monitoring concentrations evaluated from 1996 to 1998 (Dennehy et al. 1999). Contrasts of 2002 and 2003 means with those from 1996, 1997, and 2000 (Figure 2) revealed no further decreases in susceptibility. Current levels of susceptibility of Arizona whiteflies to buprofezin are within the range observed since 1997.

Fenpropathrin + Acephate (Danitol[®] + Orthene[®])

Statewide levels of resistance to synergized pyrethroid insecticides of whiteflies from Arizona cotton have declined dramatically since 1995. This is demonstrated by strikingly higher grand mean mortality observed in bioassays of fenpropathrin + acephate mixtures (Figure 3). However, the yearly percentage of individual cotton fields with $\geq 20\%$ resistant whiteflies has oscillated widely from as high as 60% to as low as 10% of locations tested (Figure 3). A concentration of 10 $\mu\text{g/ml}$ fenpropathrin mixed with 1000 $\mu\text{g/ml}$ acephate was previously shown to discriminate between whiteflies susceptible and resistant to this mixture (Dennehy and Williams 1997). Sivasupramaniam et al. (1997) subsequently demonstrated that susceptibility to fenpropathrin + acephate mixtures reflected susceptibility to all synergized pyrethroid mixtures commonly being used against whiteflies in Arizona. Field trials (Simmons and Dennehy 1996) indicated that performance of synergized pyrethroid mixtures was acceptable at locations with a frequency of $< 20\%$ resistant whiteflies.

Fifty percent of cotton fields monitored in 2002 and 25% of cotton fields monitored in 2003 had frequencies of resistance exceeding the critical frequency (Figure 4). Thus, we conclude that some producers obtained inadequate control of whiteflies from expenditures on synergized pyrethroid treatments during these years. However, our whitefly collections were made late in the season and some undoubtedly reflected susceptibility of populations after they had been treated. Synergized pyrethroids mixtures are used for controlling a number of pests of Arizona cotton, in addition to whiteflies. All such treatments, irrespective of the intended target pest, can result in increased frequencies of resistance in treated whiteflies.

Synergized pyrethroid treatments against whiteflies can be highly effective and economical and they constitute an important element of our whitefly resistance management strategy. However, if used too frequently or too early in the season, they can be detrimental to whitefly management and, therein, increase production costs. It is for these

reasons that the 1996 whitefly resistance management program for Arizona cotton recommended that synergized pyrethroids be used against whiteflies in cotton only late in the season and be limited to a maximum of two applications per season (Dennehy et al. 1996). The relatively low cost of synergized pyrethroid treatments of whiteflies provides an incentive to overuse them and/or to use them against whiteflies at times other than the late season. Without appropriate restraint, Arizona cotton producers could slip back into insecticide use patterns that triggered the resistance-related whitefly outbreaks experienced in 1994 and 1995. Thus, in extension education programs during the coming year producers will be reminded of the importance of delaying use of pyrethroids and other broadly-toxic insecticides until late in the season, as well as the value of limiting pyrethroids to no more than 2 applications per season.

Imidacloprid (Admire/Provado)

Whiteflies from cotton throughout Arizona were extremely susceptible to the neonicotinoid insecticide, imidacloprid, in 2002 and 2003, despite the fact that this insecticide has been used extensively on vegetable and melon crops since 1993 (Figure 5). Susceptibility to imidacloprid dropped strikingly in Arizona whiteflies from 1995 to 1998 (Figure 5). Bioassays of collection made in 1998 frequently had 20 to 50% survivors of 100 and 1000 µg imidacloprid/ml treatments. We now occasionally find such resistant populations in greenhouse-grown ornamentals from Arizona. However, current levels of susceptibility of field populations from throughout Arizona are so high that we rarely observe greater than 10% survivors of assays of 10 µg imidacloprid/ml. Moreover, yearly testing of whiteflies from poinsettias and other ornamentals has yielded no indications of problems with performance of neonicotinoids against whiteflies in greenhouses (data not shown). All Arizona whitefly collections evaluated in 2002 and 2003 were also highly susceptible (data not shown) to acetamiprid (Intruder[®]) and thiamethoxam (Actara[®]/Centric[®]/Platinum[®]).

SUMMARY

No major problems regarding field performance of insecticides against whiteflies were observed or reported in Arizona in 2002 or 2003. However, monitoring confirmed declining susceptibility to pyriproxyfen (Knack) and showed that whiteflies possessing reduced levels of susceptibility could be detected in all cotton-producing areas of the state. Susceptibility to buprofezin (Courier/Applaud) has not changed significantly since 1997. Mean susceptibility to synergized pyrethroids (e.g., Danitol + Orthene) has increased strikingly on a statewide basis since 1995. However, 50 and 25% of cotton fields sampled in 2002 and 2003, respectively, had resistance levels expected to result in inadequate field performance of synergized pyrethroid treatments. Whiteflies from throughout Arizona were highly susceptible to imidacloprid (Admire/Provado) and two other neonicotinoid insecticides, acetamiprid and thiamethoxam.

ACKNOWLEDGEMENTS

We thank the Arizona Cotton Growers Association and Cotton Incorporated for supporting these studies. Facilities of the Extension Arthropod Resistance Management Laboratory are provided by the University of Arizona. We thank Peter Else and the staff of the University of Arizona Campus Agricultural Center for assistance in maintaining laboratory and greenhouse spaces. Peter Ellsworth and John Palumbo provided critical leadership of the Cross-Commodity Coordinating Committee and Cotton and Vegetable IPM teams.

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Table 1a. Locations from which whiteflies were successfully collected in 2002 and brought to the EARML facilities in Tucson for rearing and testing.

Location	Host	Date	Sample ID
Calexico, CA	Melons	1-Sep	02- 28
Campus Ag Ctr, Tucson, AZ	Cotton	6-Aug	02- 18
Casa Grande, AZ	Cotton	16-Sep	02- 40
Coolidge, AZ	Cotton	2-Oct	02- 49
Eloy, AZ	Cotton	27-Jul	02- 15
Gilbert, AZ	Cotton	15-Oct	02- 100
Goodyear, AZ	Melons	10-Jul	02- 9
Harquahala Valley, AZ	Melons	24-Oct	02- 102
Maricopa Ag. Ctr., AZ, #1	Melons	10-Jul	02- 8
Maricopa Ag. Ctr., AZ, #2	Cotton	16-Sep	02- 41
N. Gila Valley, AZ	Cotton	19-Aug	02- 25
Paloma, AZ	Cotton	25-Sep	02- 47
Parker Valley, AZ	Cotton	25-Sep	02- 43
Queen Creek, AZ	Cotton	15-Oct	02- 101
S. Gila Valley, AZ	Melons	19-May	02- 1
Somerton, AZ	Melons	12-Jun	02- 7
Stanfield, AZ	Cotton	2-Oct	02- 48
Tucson Retail Nursery #5	Poinsettia	16-Dec	02- 108
Vicksburg, AZ	Cotton	24-Oct	02- 103
Yuma Valley Ag. Ctr., AZ, #1	Melons	19-May	02- 2
Yuma Valley Ag. Ctr., AZ, #2	Melons	23-Jul	02- 13
Yuma, AZ #1	Melons	12-Jun	02- 5
Yuma, AZ #2	Melons	12-Jun	02- 6

Table 1b. Locations from which whiteflies were successfully collected in 2003 and brought to the EARML facilities in Tucson for rearing and testing.

Location	Host	Date	Sample	ID
Avondale, AZ	cotton	11-Sep	03-	18
Buckeye, AZ, #1	cotton	24-Sep	03-	30
Buckeye, AZ, #2	cotton	23-Jul	03-	113
Chandler, AZ	cotton	25-Aug	03-	11
Coolidge, AZ	cotton	11-Sep	03-	19
Eloy, AZ	cotton	3-Sep	03-	114
Eloy, AZ	melons	7-Jul	03-	106
Gadsen, AZ	melons	14-Jul	03-	109
Goodyear, AZ	melons	16-Jun	03-	101
Harquahala Valley, AZ	cotton	8-Jul	03-	107
Holtville, CA	cotton	20-Aug	03-	10
Litchfield Park, AZ, #1	cabbage	8-Sep	03-	17
Litchfield Park, AZ, #2	melons	26-Jun	03-	103
Maricopa Ag. Center, AZ	cotton	21-Sep	03-	31
Mohave Valley, AZ	cotton	10-Aug	03-	3
Mohawk Valley, AZ	melons	29-Jun	03-	104
North Gila Valley, AZ	cotton	19-Aug	03-	8
Palo Verde, AZ	cotton	25-Aug	03-	12
Parker Valley, AZ #2	cotton	21-Jul	03-	112
Parker Valley, AZ, # 1	cotton	10-Aug	03-	4
Parker Valley, AZ, #3	melons	10-Aug	03-	5
Picacho, AZ	cotton	21-Sep	03-	24
Stanfield, AZ	cotton	3-Oct	03-	115
Stanfield, AZ	melons	16-Jul	03-	111
Tacna, AZ	cotton	31-Aug	03-	16
Tucson Retail Nursery #1	hibiscus	18-Jun	03-	102
Tucson Retail Nursery #1	lantana	19-Dec	03-	116
Yuma Ag. Center, AZ,	melons	29-Jun	03-	105
Yuma, AZ, #1	cotton	19-Aug	03-	7
Yuma, AZ, #2	melons	19-May	03-	100

Table 1c. Summary of bioassay methods employed for each insecticide tested against whiteflies in 2002 and 2003.

	<u>Pyriproxyfen</u>	<u>Imidacloprid</u>	<u>Fenpropathrin (+ Acephate)</u>	<u>Buprofezin</u>	<u>Thiamethoxam</u>	<u>Acetamiprid</u>
<u>Concentrations</u>	control, 0.01, 0.1, 1.0	control, 1, 10, 100, 1000	control, 10, 100 (+1000)	control, 8, 100, 1000	control, 1, 10, 100, 1000	control, 1, 10, 100, 1000
<u>Replications</u>	6 plant replicates/>20 eggs/leaf	10 vial reps, 25 adults/vial	6 vial reps, 25 adults/vial	6 plant reps, >20 nymphs/plt	6 vial reps, 25 adults/vial	6 vial reps, 25 adults/vial
<u>Method</u>	seedling in vial	Residual leaf-disc on agar	Residual leaf-disc on agar	seedling in vial	Residual leaf-disc on agar	Residual leaf-disc on agar
<u>Stage treated</u>	egg	adult	adult	N1 stage	adult	adult
<u>Treatment Method</u>	leaf-dip, 20 sec	24h systemic uptake	leaf-dip, 10 sec	leaf-dip 20 sec	leaf-dip 10 sec	leaf-dip 10 sec
<u>Duration</u>	7 days exposure	48h exposure	48h exposure	9 days exposure	48h exposure	48h exposure
<u>Notes</u>	24h ovip period, followed by 20s leaf dip, read 7 days after dipping.	Small seedling (2-4 true leaf stage), cut stem above root line. Put into imida solution for 24h.	Small seedling (2-4 true leaf stage), cut leaf discs and dip for 10 sec into solution.	24h ovip period, followed by 8 days to develop to N1, 20 sec leaf dip, read 9 days after dipping.	Small seedling (2-4 true leaf stage), cut leaf discs and dip for 10 sec into solution.	Small seedling (2-4 true leaf stage), cut leaf discs and dip for 10 sec into solution.

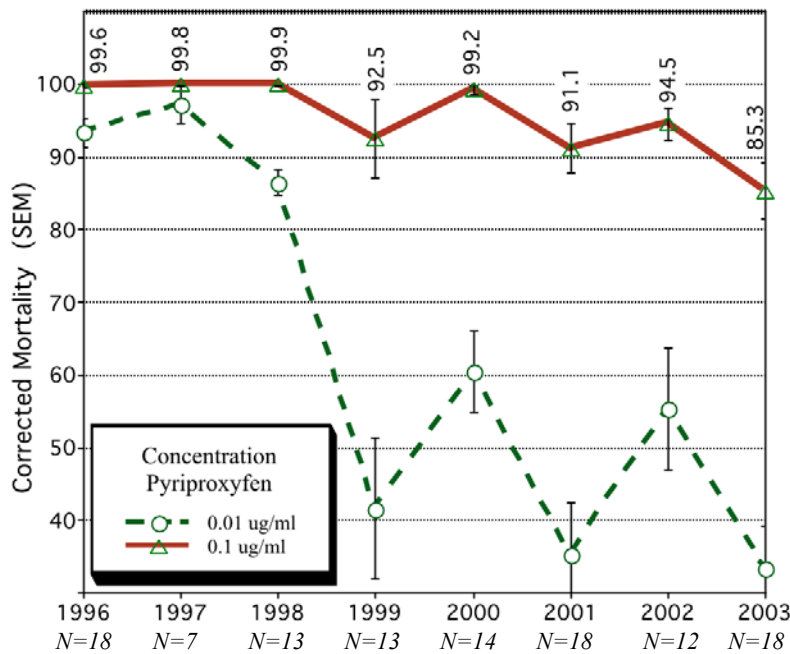


Figure 1a. Average susceptibility to pyriproxyfen (Knack[®]) of whiteflies from Arizona cotton, 1996-2003, as depicted by survivorship in bioassays of 0.01 and 0.1 μg pyriproxyfen/ml. Note that very few whiteflies survived 0.1 μg /ml bioassays during the first three years that pyriproxyfen was used in Arizona: 1996, 1997 and 1998. The number of collections tested is specified for each year.

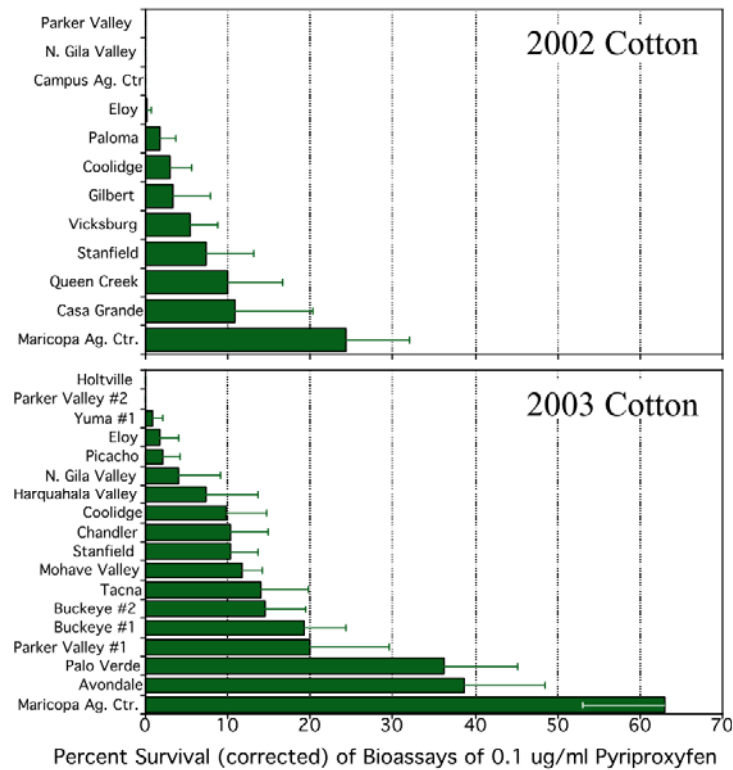


Figure 1b. Susceptibility to pyriproxyfen (Knack[®]) in 2002 and 2003 of whiteflies from cotton. Shown are the proportions of whiteflies surviving discriminating concentration bioassays of 0.1 μg pyriproxyfen/ml.

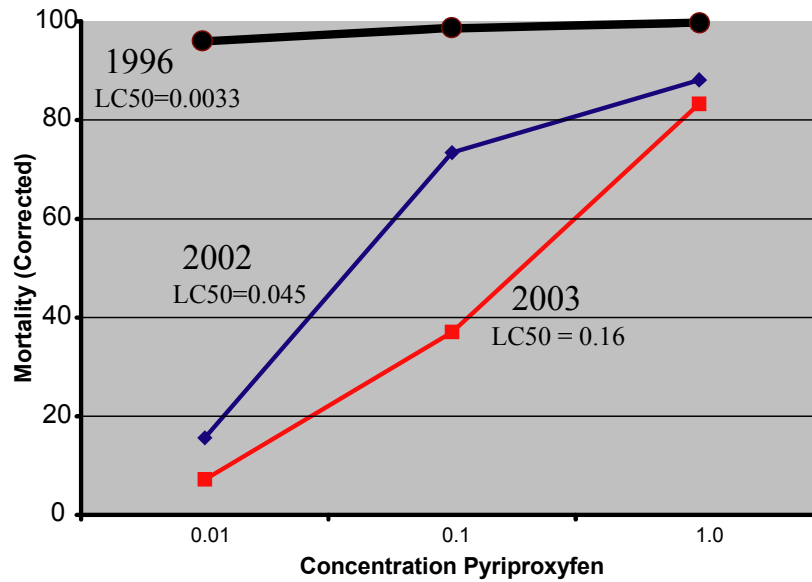


Figure 1c. Whiteflies least susceptible to pyriproxyfen (Knack[®]) in 2002 and 2003 were collected from cotton at the Maricopa Agricultural Center. Relative to 1996, the 2002 and 2003 collections were 13-fold and 48-fold less susceptible to pyriproxyfen. Some data points used to derive the probit line for 1996 collection are not shown.

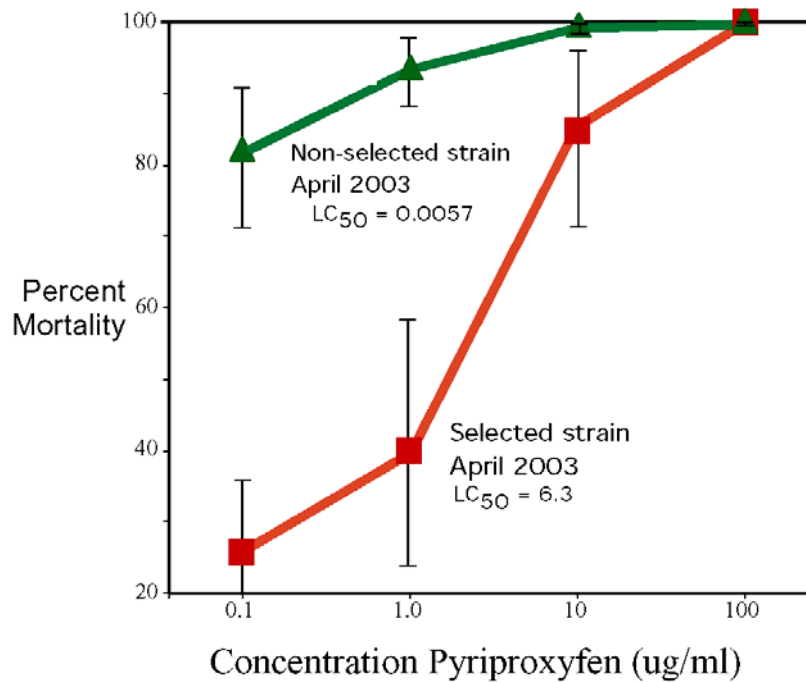


Figure 1d. Laboratory selection of the 2002 Queen Creek, Arizona, population of *Bemisia tabaci*. Two exposures to pyriproxyfen (Knack[®]) in the laboratory increased resistance levels in this strain by over 1000-fold. LC₅₀ data for the non-selected strain were generated from full response lines (data not shown).

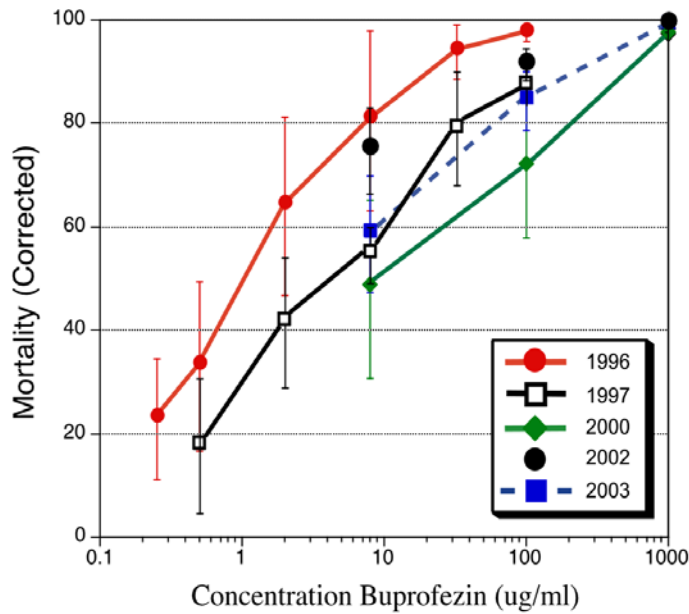


Figure 2. Grand mean corrected mortality (\pm stdev.) of whiteflies collected from Arizona cotton in 1996 through 2003 and bioassayed with buprofezin (Courier[®]/Applaud[®]). Susceptibility declined moderately from 1996 to 2000. Susceptibility in 2002 and 2003 was intermediate to this range. Numbers of collections evaluated per year were: 1996, N=9; 1997, N=7; 2000, N=14; 2002, N=12; 2003, N=15.

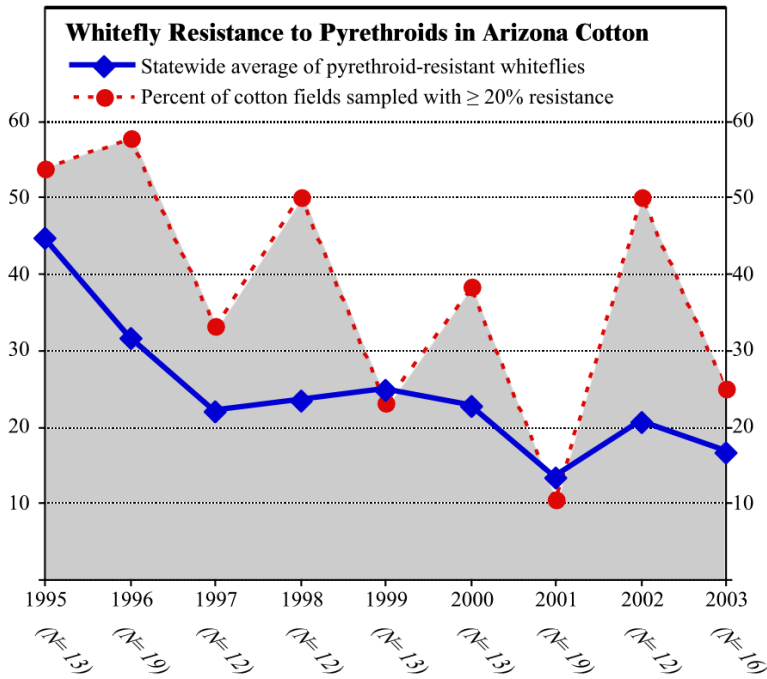


Figure 3. Resistance of Arizona whiteflies from cotton to synergized pyrethroids in 1995 to 2003, as reflected susceptibility to mixtures of fenpropathrin (Danitol[®]) + acephate (Orthene[®]). Yearly averages of the whiteflies surviving discriminating concentration bioassays (10 $\mu\text{g/ml}$ fenpropathrin + 1000 $\mu\text{g/ml}$ acephate) show overall declining levels of resistance. However, in some years more than half of Arizona fields evaluated had resistance frequencies that were too high to obtain adequate field efficacy of synergized pyrethroid mixtures.

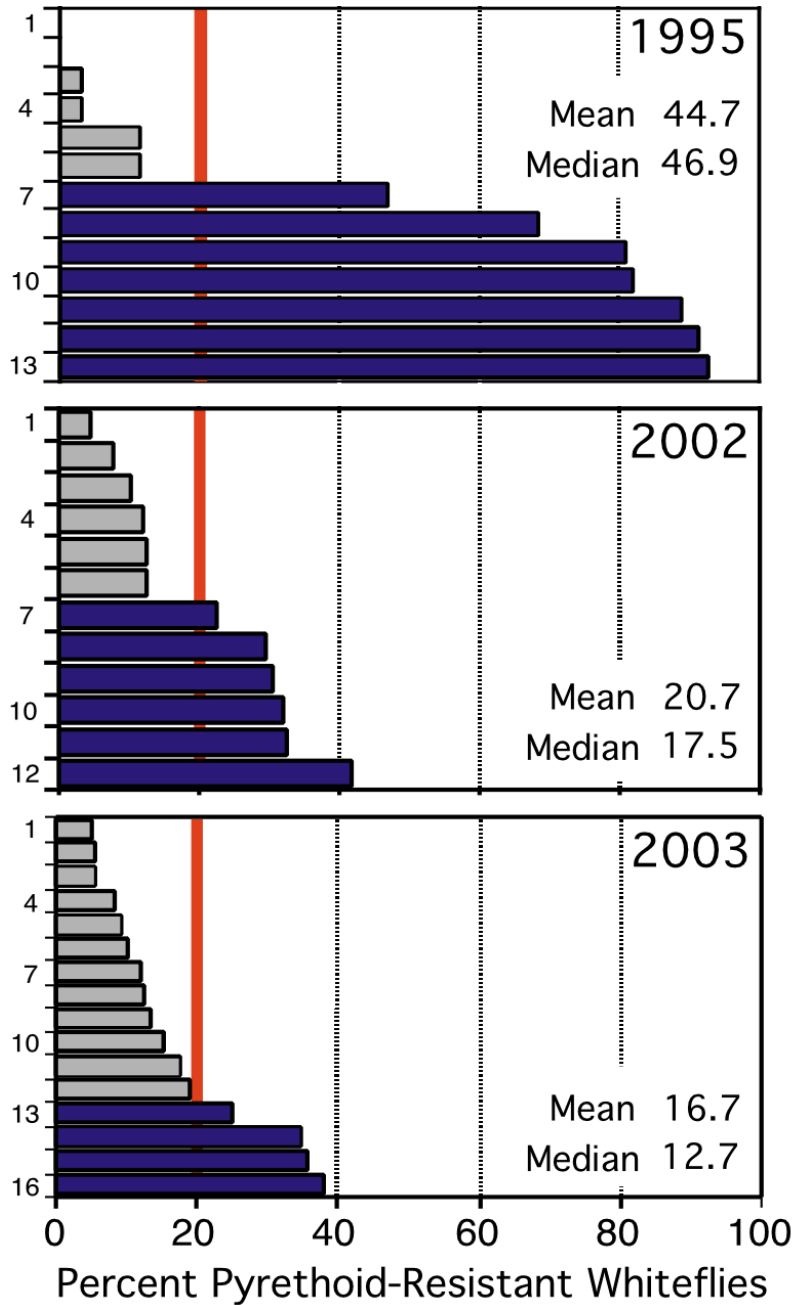


Figure 4. Susceptibility to synergized pyrethroid insecticides of Arizona whiteflies collected from cotton in 2002 (12 collections) and 2003 (16 collections), relative to 1995 (13 collections). Shown is the percentage of whiteflies from each sample surviving a discriminating concentration of 10 µg/ml fenpropathrin (Danitol®)+ 1000 µg/ml acephate (Orthene®). This concentration has been shown to kill susceptible whiteflies. The vertical line at 20% indicates the critical frequency above which resistance demonstrably impairs field performance.

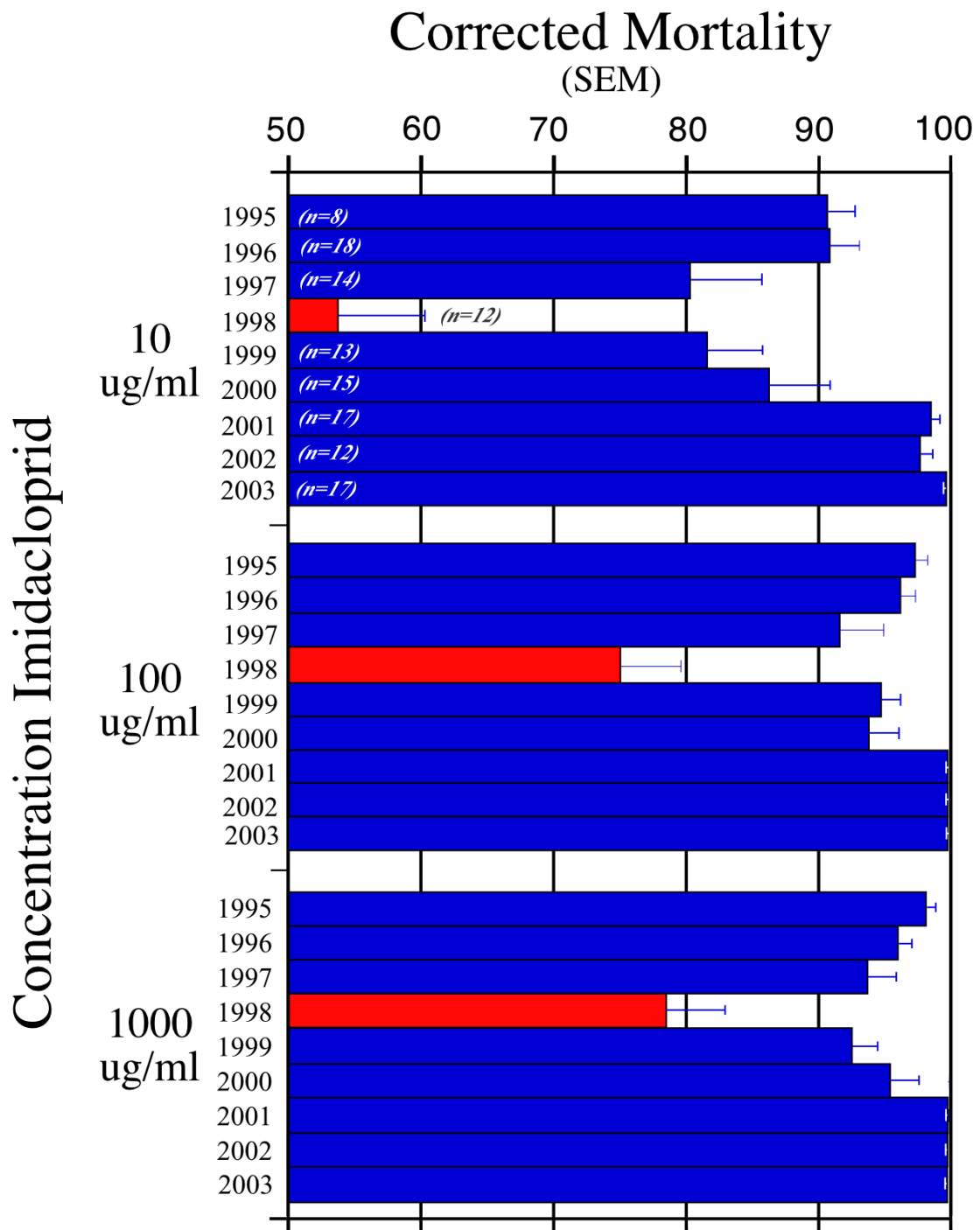


Figure 5. Susceptibility to imidacloprid (Admire®/Provado®) of Arizona whiteflies collected from cotton, 1995-2002. Susceptibility declined sharply from 1995 to 1998 but was fully regained in subsequent years.